

β-半乳糖苷酶染色实验报告

一、实验器材及试剂

1、 实验器材

| 名称 | 厂家 | 型号 |
|---------|------------|--------------------|
| 冰冻切片机 | Thermo | CRYOSTAR NX50 |
| 切片刀 | 上海徕卡仪器有限公司 | LEICA 819 |
| 载玻片 | Wanwu | |
| 正置光学显微镜 | 日本尼康 | NIKON ECLIPSE E100 |
| 成像系统 | 日本尼康 | NIKON DS-U3 |

2、 主要实验试剂

| 试剂名称 | 厂家 | 货号 |
|--------------|--------|-------|
| OCT 包埋剂 | Sakura | 4583 |
| β-半乳糖苷酶染色液 A | Wanwu | G1073 |
| β-半乳糖苷酶染色液 B | Wanwu | G1073 |
| β-半乳糖苷酶染色液 C | Wanwu | G1073 |
| β-半乳糖苷酶染色液 D | Wanwu | G1073 |
| β-半乳糖苷酶染色液 E | Wanwu | G1073 |
| β-半乳糖苷酶染色液 F | Wanwu | G1073 |
| 甘油明胶封片剂 | Wanwu | G1402 |

二、染色步骤

- 1、工作液配制：**染色液 A 940ul，染色液 B 10ul，β-半乳糖苷酶染色液 D50ul，依次加入，混匀即为工作液。
- 2、固定：**取一张没经过固定的新鲜冰冻切片自然晾干，β-半乳糖苷酶染色液E固定15min，倾去固定液，蒸馏水洗3遍；
- 3、β-半乳糖苷酶染色：**甩干片子上的水，将工作液滴于组织上，于37°C孵育16至18h。
- 4、**倾去工作液，蒸馏充分水洗3次。



5、 β -半乳糖苷酶染色液F孵育3min后，水洗。

6、**脱水封片**：切片依次放入无水乙醇I 5min -无水乙醇II 5min-无水乙醇III5min -二甲苯I5min-二甲苯II5min透明，中性树胶封片。

三、结果判读：

衰老细胞呈蓝色。

四、注意事项：

- 1、标本需要是新鲜的冰冻切片或细胞，不能经过固定。
- 2、染液需要在-20°C的条件下储存，保质期一年。
- 3、工作液要现配现用，不可保存。

β -galactosidase Staining Experiment Report

I. Experimental equipment and reagents

1. Experimental equipment

| Equipment name | Manufacturer | Model No. |
|-----------------------------|-------------------------------------|--------------------|
| Freezing microtome | Thermo | CRYOSTAR NX50 |
| Slicing knife | Shanghai Leica Instrument Co., Ltd. | LEICA 819 |
| Microscope slides | Wanwu | |
| Upright electron microscope | Japan NIKON | NIKON ECLIPSE E100 |
| Imaging system | Japan NIKON | NIKON DS-U3 |

2. Main experimental reagents

| Reagent name | Manufacturer | Item No. |
|--|--------------|----------|
| OCT Embedding medium | Sakura | 4583 |
| β -galactosidase staining solution A | Wanwu | G1073 |
| β -galactosidase staining solution B | Wanwu | G1073 |
| β -galactosidase staining solution C | Wanwu | G1073 |
| β -galactosidase staining solution D | Wanwu | G1073 |
| β -galactosidase staining solution E | Wanwu | G1073 |
| β -galactosidase staining solution F | Wanwu | G1073 |
| Glycerin Jelly | Wanwu | G1402 |

II. Staining steps

1. Preparation of working solution: 940ul of staining solution A, 10ul of staining solution B, and 50ul of β -galactosidase staining solution D, add them in sequence and mix well, then the working

solution is made.

2. Fixation: Take a fresh frozen section that has not been fixed and let it dry naturally, fix it with β -galactosidase staining solution E for 15 min, pour out the fixation solution, and wash it three times with distilled water.

3. β -galactosidase staining: Spin off the water on the section, drop the working solution onto the tissue, and incubate at 37°C for 16 to 18 hours.

4. Pour out the working solution and wash adequately with distilled water for 3 times.

5. After incubation with β -galactosidase stain F for 3 min, wash with water.

6. Dehydration and sealing: Put the sections in sequence into Anhydrous ethanol I for 5min - Anhydrous ethanol II for 5min - Anhydrous ethanol III for 5min - Xylene I for 5min - Xylene II for 5min for transparency, seal with neutral gum lastly.

III. Interpretation of results:

Senescent cells are blue.

IV. Precautions:

1. Specimens need to be fresh frozen sections or cells, and can't be fixed.
2. The staining solutions need to be stored at -20°C, and the shelf life is one year.
3. The working solution should be prepared freshly and use immediately, and cannot be stored.